DATA SHEET-V1 REMBRANDT® CEP1-FISH DETECTION

Intended use

- The REMBRANDT® CEP1-FISH assay is an invitro diagnostics medical device intended for the detection of the human centromeric region of chromosome 1 by means of in situ hybridization.
- II. The REMBRANDT® CEP1-FISH assay is intended for the detection of the centromeric region of chromosome 1 in fixed cells. The clinical interpretation of the results should not be established on the basis of a single test result. A precise diagnosis, in fact, should take into consideration clinical history, symptoms, as well as other possible test data.
- III. The REMBRANDT® CEP1-FISH assay kit is a quantitative assay for the detection of the centromeric region of the human chromosome 1.
- IV. The intended users are qualified laboratory employees in cytology and/or pathology. The product is intended for professional use.

Clinical relevance

Within the chromosomes, the satellite DNA forms essential structures of chromosomes, i.e. telomeres and centromeres, ensuring protection and stability of chromatin of these sites. Satellites are highly variable in their sequence and in the number of copies both within a species and between closely related species that indicates rapid evolution of satellites. Satellite repeats are mostly located in centromeric and pericentromeric regions of chromosomes (Shatskikh et al., 2020). Errors within the pathways that regulate the assembly of the centromeres, can result in mis-segregation and aneuploidy of the chromosomal copy numbers. These are features for cancerous cells. Besides this, it can also result in chromosomal instability and, eventually chromosomal breakage (Barra & Fachinetti, 2018).

Probe specification

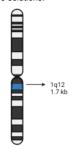
The CEP1-FISH probe consists of a 1.7 kb probe and is available in two colours. For an orange detection the AF555 assay is available and for green detection the AF488 assay is available. The CEP1-FISH probe is able to

bind to the satellite repeats in the 1q12 region heterochromatin region of chromosome 1. The CEP1-FISH probes are pre-mixed in a hybridization mixture





(formamide, dextran sulphate and SSC) and are ready to use solutions.



Test principle

In a fluorescent in situ hybridization assay, a double stranded DNA probe labelled with a fluorochrome is used. The labelled DNA probe is diluted in a hybridization mixture. The hybridization mixture containing the DNA probe is added to the specimen. A co-denaturation will ensure that the genomic DNA and the probe DNA become single stranded. After denaturation, the probe DNA is able to hybridize to its complementary target sequences in the cells. In the REMBRANDT® CEP1-FISH assay, the fluorochrome is attached to the probe and the signals can be visualized directly by fluorescent microscopy after hybridization.

Reagents provided

Product na Labelled Cl • CEP probe detection		Product number depending on label and s C701P.2000.05	Amount size choice)
CEP probe detection	1-FISH green	C701P.2000.10	∑ 10 T
CEP probe detection	1-FISH orange	C701P.3000.05	∑ 5 T
CEP probe detection	1-FISH orange	C701P.3000.10	∑ 10 T
Pepsin pow Pepsin dilu PanWash SSC	ent	R011R.0000 R018R.0000 R025R.0000	1 g 15 ml 15 ml
Fluorescen Mounting n	-	Z000R.0050	1 ml

Assay procedure

REMBRANDT® CEP1-FISH detection assay procedure for cytological specimen.

- Incubate slides in pre-heated proteolytic work solution (prepare according to section 1.9 of Manual-FISH) (R011R.000 + R018R.000) at 37 °C (100 µg/ml) for 15 minutes followed by a brief rinsing in 0.01M HCI (1x 2 minutes) and subsequent rinses in PBS (2x 1 minute)
- II. Flush wash slides in deionised water, followed by dehydration in graded ethanol series (ethanol 70%, 96%, 96%, 100%, 100%) 1 minute each and air-dry slides for 15 minutes

Do not treat more than 5 slides at the same time, because the temperature of the pre-heated solutions may drop dramatically, thus causing incomplete pre-treatment. Additionally, allow the slides to air-dry as recommended; otherwise sections will be lost.

III. Homogenize probe solution (C701P.XXXX.YY) and spin briefly. Apply 15 µI of probe solution to each specimen. Cover all specimens with a cover slip (avoid air bubbles). Denature slides on a 80 °C hotplate or other heating device for 3 minutes

Work in a pre-set order to ensure that all slides have been incubated at 80 °C for the exact same time. Do not denature more than 5 slides at the same time, the temperature of the heating device may drop dramatically, thus causing incomplete denaturation.

- IV. Transfer the slides into a moist environment and incubate for 16 hours at 37 °C.
- V. Remove coverslips by soaking the slides in PBS at room temperature
- VI. Incubate the slides in diluted, pre-heated PanWash 4 (R025R.000) (prepare according to section 1.9 of Manual-FISH)

Do not incubate more than 5 slides at the same time in PanWash 4, the temperature of the PanWash 4 may drop dramatically, causing wrong stringency conditions.

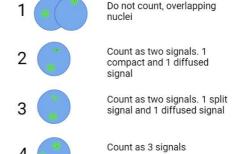
- VII. Incubate the slides in PBS at room temperature for 1 minute
- VIII. Dehydrate the slides in graded ethanol series (70%, 96%, 96%, 100%, 100%) 1 minute each and air-dry the slides for 15 minutes (in the dark)
- IX. Mount the slides by applying mounting medium (Z000R.0050) and coverslip

Interpretation of results

Hybridization of the CEP1-FISH probe is viewed using a fluorescence microscope equipped with appropriate excitation and emission filters. For green detection: λ_{exc} 492 nm, λ_{em} 517 nm and for orange detection: λ_{exc} 555 nm, λ_{em} 572 nm. Allowing visualization of both the intense green or orange fluorescent signal concentrated at the centromeric region of chromosome 1 and the blue counterstained chromosomes and nuclei enumeration of chromosome 1 is conducted by microscopic examination of interphase nuclei. The fluorescently-stained centromeres of chromosome 1 stand out brightly against the general fluorescence of the nucleus. The CEP1 procedure enables visual enumeration e.a. monosomy disomy, or trisomy of the centromeric region of chromosome 1 within the nuclei. The assay results are reported as the percentage of nuclei with 0, 1, 2, 3, 4, and >4 fluorescent signals. Each fluorescent signal corresponds to the centromere of chromosome 1.

Enumerate the fluorescent signals in the interphase nucleus using a 40X or 63X magnification. Objectives with higher magnification (eg, 63X or 100X) should be used to verify or resolve questions about split or diffused signals. Enumerate at least 100 nuclei per slide for accurate analysis.

- •Two signals in close proximity and approximately the same sizes but not connected by a visible link are counted as 2 signals.
- •Count a diffuse signal as 1 signal if diffusion of the signal is contiguous and within an acceptable boundary.
- •Two small signals connected by a visible link are counted as 1 signal.
- •Enumerate the number of nuclei with 0, 1, 2, 3, 4, or >4 signals. Count nuclei with 0 signals only if there are other nuclei with at least 1 signal present in the field of view. If the accuracy of the enumeration is in doubt, repeat the enumeration in another area of the slide.
- •Do not enumerate nuclei with uncertain signals



Performance characteristics

Analytical Sensitivity and Specificity

The analytical sensitivity and specificity were investigated within PanPaths analytical performance assessment. Precision was investigated for the CEP1-FISH assay and results are available upon request.

Analytical sensitivity

The analytical sensitivity was determined in three levels. The normal cut-off percentage was determined based on assessment of 200 individual nuclei in two samples of healthy interphase lymphocytes from peripheral blood. The beta inversion was used to determined the percentage of normal-cut off. For the noise-to-signal percentage, the noise and signal values were determined for 100 signals in interphase lymphocytes from peripheral blood of two independent samples. The differences between noise and signal were evaluated using a 95% confidence interval. For the hybridization efficiency, 200 individual nuclei were assessed for the presence of FISH signals.

Performance characteristic Normal cut-off percentage	Outcome 8.6%
Noise-to-signal cut-off	4.8%
percentage Hybridization efficiency	100%

Analytical specificity

The analytical specificity was determined in two levels. The theoretical specificity was determined by sequencing analysis of the probe DNA and mapping on Hg38. The practical specificity was determined by assessing the hybridization pattern in metaphase chromosomes of interphase lymphocytes from peripheral blood.

Performance characteristic	Outcome	
Theoretical specificity	Mapped on chromosome q12	1,
Practical specificity	100%	

Clinical performance

The clinical performance was not determined for the REMBRANDT® CEP1-FISH assays since the assays do not detect a specific condition. The clinical performance is demonstrated by scientific validity studies.

Limitations of Procedure

- i) The CEP1-FISH assay is solely applicable for the detection of the centromeric region of the human chromosome 1, which may be present in cell preparations (cytological specimen i.e. interphase lymphocytes from peripheral blood samples).
- ii) Either human tissue sections or human cytological preparations may be used. Samples must be fixed in buffered formalin or alcohol. In tissue sections are required, the sections should be prepared in a 4 µm

thickness. Furthermore, the tissues should be glued to the glass slides with a bio-adhesive (e.g. organ silane), dried at room temperature, subsequently dried at 37 °C overnight and lastly completely deparaffinized in xylene and alcohol series and air dried.

- iii) Cytological specimen should be prepared as required by the user, fixed with cytological fixation agent, rinsed in distilled water prior to the ISH procedure and air dried
- iv) Many factors can influence the performance of the ISH procedure. Failure in detection can be due to i.e. improper sampling, handling, the time lapse between tissue removal and fixation, the size of the tissue specimen in the fixation medium, the fixation time, processing fixed tissue, the thickness of the section, the bio-adhesive on the slide, deparaffinisation procedure, incubation times, proteolytic pre-treatment, detection reagents, incubation temperatures and interpretation of results
- v) The performance of the ISH procedure is also affected by the sensitivity of the method and the presence of the centromeric region of chromosome 1. In case the limit of the sensitivity is reached a false negative reaction may be the result.
- vi) The CEP1-FISH assay results should not be relied on in case the sampling, sampling method, sample quality, sample preparation, reagents used, controls and procedure followed are not optimal or as described the working protocols.
- vii) The clinical interpretation of the results should not be established on the basis of a single test result. A precise diagnosis should take into consideration clinical history as well as data obtained from other molecular test (i.e. WGS).
- viii) Therapeutic considerations based on the result of this test alone should not been taken. Results should be verified by other traditional diagnostic methods such as but not limited to clinical history, symptoms, as well as morphological data.
- ix) The medical profession should be aware of risks and factors influencing the fluorescent signal intensity while interpretating the test result. Microscopy settings might influence the signal intensity and/or interpretation.
- x) Laboratory personnel performing the test should be trained and knowledgeable to be able to interpret the test results.

Storage and handling

Store kit and its contents at 2-8°C. Store the dissolved and aliquoted reagents at recommended temperatures. When used and stored as indicated, the kit is stable until the expiry date printed on the box.

Produc	t	Product number	Storage conditions
CEP1	Probe	C701P.XXXX	2-8 °C
orange detectio	or green n		

Pepsin powder	R011R.0000	Powder: 2-25°C, ambient temperature
Pepsin diluent	R018R.0000	Dissolved: -20°C Concentrated solution and diluted: 2-25°C, ambient
PanWash 4, 25X SSC	R025R.0000	temperature Concentrated solution and diluted: 2-25°C, ambient temperature
Fluorescent mounting medium	Z000R.0050	2-8 °C



Hazard statements

H315 - Causes skin irritation
H319 - Causes serious eye irritation
H351 - Suspected of causing cancer
H360D - May damage the unborn child
H373 - May cause damage to organs through prolonged or repeated exposure

Precautionary Statements

P202 - Do not handle until all safety precautions have been read and understood

P280 - Wear protective gloves/protective clothing/eye protection/face protection

P302 + P352 - İF ON SKIN: Wash with plenty of water and soap P305 + P351 + P338 - IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing

P308 + P313 - IF exposed or concerned: Get medical advice/attention

P362 + P364 - Take off contaminated clothing and wash it before reuse

P405 - Store locked up

Additional information

Product in combination with other devices

The REMBRANDT® DNA probes are intended for standalone usage. The in vitro diagnostic is intended to be used in combination with standard cytological preparation methods, hot plate(s), stove(s), incubation device(s), water bath(s), temperature and incubation time control(s), proteolytic-, detection- and other reagents (not supplied with this reagent) and a microscope. The combination has been tested and validated. Since the standard cytological preparation methods, hot plate(s), stove(s), incubation device(s), water bath(s), temperature controls, incubation time control(s) and other not supplied reagents such as but not limited to proteolytic reagents, detection reagents and a microscope is not combined with the device as a

product, conformity with the essential requirements is not applicable. Assay validation criteria are mentioned in 'Interpretation of the Results' and are also depending on clinical state of the sample, which may influence the validation criteria.

For additional information regarding the REMBRANDT® assays, a manual is included which specifies the following subjects:

- Controls
- Materials required but not included
 - Storage and shelf-life
- Performance precautions
- Preparations of reagents
- Specimen collection
- Quality control
- Trouble shooting guide

Technical assistance

For technical assistance regarding the products performance, please contact info@panpath.nl or call +31 495499090. Visit our website for reprints of datasheets or additional documentation. www.panpath.nl

Literature list

Barra, V., & Fachinetti, D. (2018). The dark side of centromeres: types, causes and consequences of structural abnormalities implicating centromeric DNA. *Nature Communications*, 9(1). https://doi.org/10.1038/s41467-018-06545-y

Shatskikh, A. S., Kotov, A. A., Adashev, V. E., Bazylev, S. S., & Olenina, L. V. (2020). Functional Significance of Satellite DNAs: Insights From Drosophila. Frontiers in Cell and Developmental Biology, 8(May), 1–19. https://doi.org/10.3389/fcell.2020.00312

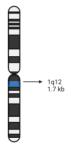
DATA SHEET-V1 REMBRANDT® CEP1-FISH DETECTION RESEARCH USE ONLY (RUO)

RUO



Ref	C701K.2000.05	\sum	5 T
	C701K.2000.10	$\overline{\Sigma}$	10 T
	C701K.3000.05	Ž	5 T
	C701K.3000.10	$\overline{\Sigma}$	10 T

(formamide, dextran sulphate and SSC) and are ready to use solutions.



Intended use

- The REMBRANDT® CEP1-FISH assay is for research use only and is intended for the detection of the human centromeric region of chromosome 1 by means of in situ hybridization.
- II. The REMBRANDT® CEP1-FISH assay is intended for the detection of the centromeric region of chromosome 1 in fixed cells. A clinical diagnosis should not be established based on the performance of this test.
- III. The REMBRANDT® CEP1-FISH assay kit is a quantitative assay for the detection of the centromeric region of the human chromosome 1.
- IV. The intended users are qualified laboratory employees in cytology and/or pathology. The product is intended for professional use.

Clinical relevance

Within the chromosomes, the satellite DNA forms essential structures of chromosomes, i.e. telomeres and centromeres, ensuring protection and stability of chromatin of these sites. Satellites are highly variable in their sequence and in the number of copies both within a species and between closely related species that indicates rapid evolution of satellites. Satellite repeats are mostly located in centromeric and pericentromeric regions of chromosomes (Shatskikh et al., 2020). Errors within the pathways that regulate the assembly of the centromeres, can result in mis-segregation and aneuploidy of the chromosomal copy numbers. These are features for cancerous cells. Besides this, it can also result in chromosomal instability and, eventually chromosomal breakage (Barra & Fachinetti. 2018).

Probe specification

The CEP1-FISH probe consists of a 1.7 kb probe and is available in two colours. For an orange detection the AF555 assay is available and for green detection the AF488 assay is available. The CEP1-FISH probe is able to

bind to the satellite repeats in the 1q12 region heterochromatin region of chromosome 1. The CEP1-FISH probes are pre-mixed in a hybridization mixture

Test principle

In a fluorescent in situ hybridization assay, a double stranded DNA probe labelled with a fluorochrome is used. The labelled DNA probe is diluted in a hybridization mixture. The hybridization mixture containing the DNA probe is added to the specimen. A co-denaturation will ensure that the genomic DNA and the probe DNA become single stranded. After denaturation, the probe DNA is able to hybridize to its complementary target sequences in the cells. In the REMBRANDT® CEP1-FISH assay, the fluorochrome is attached to the probe and the signals can be visualized directly by fluorescent microscopy after hybridization.

Reagents provided

Product n		Product number	Amount
		depending on label and	size choice)
CEP	1-FISH	C701P.2000.05	Σ/
probe	green		V 51
detection			
◆CEP	1-FISH	C701P.2000.10	Σ/ _{10.T}
probe	green		V 10 I
detection			
◆CEP	1-FISH	C701P.3000.05	∑ 5 T
probe	orange		V 5 I
detection			
CEP	1-FISH	C701P.3000.10	∑ 10 T
probe	orange		V 10 I
detection			
Pepsin pov		R011R.0000	1 g
Pepsin dilu		R018R.0000	15 ml
PanWash	4, 25X	R025R.0000	15 ml
SSC			
Fluorescer		Z000R.0050	1 ml
Mounting n	nedium		

Assay procedure

REMBRANDT® CEP1-FISH detection assay procedure for cytological specimen.

- Incubate slides in pre-heated proteolytic work solution (prepare according to section 1.9 of Manual-FISH) (R011R.000 + R018R.000) at 37 °C (100 µg/ml) for 15 minutes followed by a brief rinsing in 0.01M HCI (1x 2 minutes) and subsequent rinses in PBS (2x 1 minute)
- II. Flush wash slides in deionised water, followed by dehydration in graded ethanol series (ethanol 70%, 96%, 96%, 100%, 100%) 1 minute each and air-dry slides for 15 minutes

Do not treat more than 5 slides at the same time, because the temperature of the pre-heated solutions may drop dramatically, thus causing incomplete pre-treatment. Additionally, allow the slides to air-dry as recommended: otherwise sections will be lost.

III. Homogenize probe solution (C701P.XXXX.YY) and spin briefly. Apply 15 µI of probe solution to each specimen. Cover all specimens with a cover slip (avoid air bubbles). Denature slides on a 80 °C hotplate or other heating device for 3 minutes

Work in a pre-set order to ensure that all slides have been incubated at 80 °C for the exact same time. Do not denature more than 5 slides at the same time, the temperature of the heating device may drop dramatically, thus causing incomplete denaturation.

- IV. Transfer the slides into a moist environment and incubate for 16 hours at 37 °C
- V. Remove coverslips by soaking the slides in PBS at room temperature
- VI. Incubate the slides in diluted, pre-heated PanWash 4 (R025R.000) (prepare according to section 1.9 of Manual-FISH)

Do not incubate more than 5 slides at the same time in PanWash 4, the temperature of the PanWash 4 may drop dramatically, causing wrong stringency conditions.

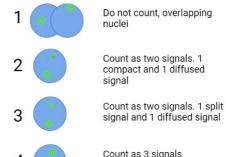
- VII. Incubate the slides in PBS at room temperature for 1 minute
- VIII. Dehydrate the slides in graded ethanol series (70%, 96%, 96%, 100%, 100%) 1 minute each and air-dry the slides for 15 minutes (in the dark)
- IX. Mount the slides by applying mounting medium (Z000R.0050) and coverslip

Interpretation of results

Hybridization of the CEP1-FISH probe is viewed using a fluorescence microscope equipped with appropriate excitation and emission filters. For green detection: $\lambda_{\text{exc}} \, 492\,$ nm, $\, \lambda_{\text{em}} \, 517\,$ nm and for orange detection: λ_{exc} 555 nm, λ_{em} 572 nm. Allowing visualization of both the intense green or orange fluorescent signal concentrated at the centromeric region of chromosome 1 and the blue counterstained chromosomes and nuclei enumeration of chromosome 1 is conducted by microscopic examination of interphase nuclei. The fluorescently-stained centromeres of chromosome 1 stand out brightly against the general fluorescence of the nucleus. The CEP1 procedure enables visual enumeration e.g. monosomy disomy, or trisomy of the centromeric region of chromosome 1 within the nuclei. The assay results are reported as the percentage of nuclei with 0, 1, 2, 3, 4, and >4 fluorescent signals. Each fluorescent signal corresponds to the centromere of chromosome 1.

Enumerate the fluorescent signals in the interphase nucleus using a 40X or 63X magnification. Objectives with higher magnification (eg, 63X or 100X) should be used to verify or resolve questions about split or diffused signals. Enumerate at least 100 nuclei per slide for accurate analysis.

- •Two signals in close proximity and approximately the same sizes but not connected by a visible link are counted as 2 signals.
- •Count a diffuse signal as 1 signal if diffusion of the signal is contiguous and within an acceptable boundary.
- •Two small signals connected by a visible link are counted as 1 signal.
- •Enumerate the number of nuclei with 0, 1, 2, 3, 4, or >4 signals. Count nuclei with 0 signals only if there are other nuclei with at least 1 signal present in the field of view. If the accuracy of the enumeration is in doubt, repeat the enumeration in another area of the slide.
- •Do not enumerate nuclei with uncertain signals



Performance characteristics

Analytical Sensitivity and Specificity

The analytical sensitivity and specificity were investigated within PanPaths analytical performance assessment. Precision was investigated for the CEP1-FISH assay and results are available upon request.

Analytical sensitivity

The analytical sensitivity was determined in three levels. The normal cut-off percentage was determined based on assessment of 200 individual nuclei in two samples of healthy interphase lymphocytes from peripheral blood. The beta inversion was used to determined the percentage of normal-cut off. For the noise-to-signal percentage, the noise and signal values were determined for 100 signals in interphase lymphocytes from peripheral blood of two independent samples. The differences between noise and signal were evaluated using a 95% confidence interval. For the hybridization efficiency, 200 individual nuclei were assessed for the presence of FISH signals.

Performance characteristic	Outcome
Normal cut-off percentage	8.6%
Noise-to-signal cut-off percentage	4.8%
Hybridization efficiency	100%

Analytical specificity

The analytical specificity was determined in two levels. The theoretical specificity was determined by sequencing analysis of the probe DNA and mapping on Hg38. The practical specificity was determined by assessing the hybridization pattern in metaphase chromosomes of interphase lymphocytes from peripheral blood.

Performance characteristic	Outcome
Theoretical specificity	Mapped on chromosome 1,
	q12
Practical specificity	100%

Limitations of Procedure

- The CEP1-FISH assay is solely applicable for the detection of the centromeric region of the human chromosome 1, which may be present in cell preparations (cytological specimen i.e. interphase lymphocytes from peripheral blood samples).
- ii) Either human tissue sections or human cytological preparations may be used. Samples must be fixed in buffered formalin or alcohol. In tissue sections are required, the sections should be prepared in a 4 µm thickness. Furthermore, the tissues should be glued to the glass slides with a bio-adhesive (e.g. organ silane), dried at room temperature, subsequently dried at 37 °C overnight and lastly completely deparaffinized in xylene and alcohol series and air dried
- iii) Cytological specimen should be prepared as required by the user, fixed with cytological fixation agent.

rinsed in distilled water prior to the ISH procedure and air dried

- iv) Many factors can influence the performance of the ISH procedure. Failure in detection can be due to i.e. improper sampling, handling, the time lapse between tissue removal and fixation, the size of the tissue specimen in the fixation medium, the fixation time, processing fixed tissue, the thickness of the section, the bio-adhesive on the slide, deparaffinisation procedure, incubation times, proteolytic pre-treatment, detection reagents, incubation temperatures and interpretation of results.
- v) The performance of the ISH procedure is also affected by the sensitivity of the method and the presence of the centromeric region of chromosome 1. In case the limit of the sensitivity is reached a false negative reaction may be the result.
- vi) The CEP1-FISH assay results should not be relied on in case the sampling, sampling method, sample quality, sample preparation, reagents used, controls and procedure followed are not optimal or as described the working protocols.
- vii) The medical profession should be aware of risks and factors influencing the fluorescent signal intensity while interpretating the test result. Microscopy settings might influence the signal intensity and/or interpretation.
- viii) Laboratory personnel performing the test should be trained and knowledgeable to be able to interpret the test results

Storage and handling

Store kit and its contents at 2-8°C. Store the dissolved and aliquoted reagents at recommended temperatures. When used and stored as indicated, the kit is stable until the expiry date printed on the box.

Product CEP1 Probe	Product number C701P.XXXX	Storage conditions 2-8 °C
orange or green detection Pepsin powder	R011R.0000	Powder: 2-25°C, ambient temperature
Pepsin diluent	R018R.0000	Dissolved: -20°C Concentrated solution and diluted: 2-25°C,
PanWash 4, 25X SSC	R025R.0000	ambient temperature Concentrated solution and diluted: 2-25°C,
Fluorescent mounting medium	Z000R.0050	ambient temperature 2-8 °C



Hazard statements

H315 - Causes skin irritation

H319 - Causes serious eve irritation

H351 - Suspected of causing cancer

H360D - May damage the unborn child

H373 - May cause damage to organs through prolonged or

repeated exposure

Precautionary Statements

P202 - Do not handle until all safety precautions have been read and understood

P280 - Wear protective gloves/protective clothing/eye protection/face protection

P302 + P352 - IF ON SKIN: Wash with plenty of water and soap P305 + P351 + P338 - IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing

P308 + P313 - IF exposed or concerned: Get medical advice/attention

P362 + P364 - Take off contaminated clothing and wash it before reuse

P405 - Store locked up

Additional information

Product in combination with other devices

The REMBRANDT® DNA probes are intended for standalone usage. The in vitro diagnostic is intended to be used in combination with standard cytological preparation methods, hot plate(s), stove(s), incubation device(s), water bath(s), temperature and incubation time control(s), proteolytic-, detection- and other reagents (not supplied with this reagent) and a microscope. The combination has been tested and validated. Since the standard cytological preparation methods, hot plate(s), stove(s), incubation device(s), water bath(s), temperature controls, incubation time control(s) and other not supplied reagents such as but not limited to proteolytic reagents, detection reagents and a microscope is not combined with the device as a product, conformity with the essential requirements is not applicable. Assay validation criteria are mentioned in 'Interpretation of the Results' and are also depending on clinical state of the sample, which may influence the validation criteria

For additional information regarding the REMBRANDT® assays, a manual is included which specifies the following subjects:

- Controls
- Materials required but not included
- Storage and shelf-life
- Performance precautions
- Preparations of reagents
- Specimen collection
- Quality control
- Trouble shooting guide

Technical assistance

For technical assistance regarding the products performance, please contact info@panpath.nl or call +31 495499090. Visit our website for reprints of datasheets or additional documentation, www.panpath.nl

Literature list

Barra, V., & Fachinetti, D. (2018). The dark side of centromeres: types, causes and consequences of structural abnormalities implicating centromeric DNA. *Nature Communications*, 9(1). https://doi.org/10.1038/s41467-018-06545-v

Shatskikh, A. S., Kotov, A. A., Adashev, V. E., Bazylev, S. S., & Olenina, L. V. (2020). Functional Significance of Satellite DNAs: Insights From Drosophila. Frontiers in Cell and Developmental Biology, 8(May), 1–19. https://doi.org/10.3389/fcell.2020.00312

DATA SHEET-V1 REMBRANDT® CEP1-ISH DETECTION RESEARCH USE ONLY (RUO)

Intended use

- I. The REMBRANDT® CEP1-ISH assay is for research use only and is intended for the detection of the human centromeric region of chromosome 1 by means of in situ hybridization.
- II. The REMBRANDT® CEP1-ISH assay is intended for the detection of the centromeric region of chromosome 1 in fixed cells. A clinical diagnosis should not be established based on the performance of this test.
- III. The REMBRANDT® CEP1-ISH assay kit is a quantitative assay for the detection of the centromeric region of the human chromosome 1.
- IV. The intended users are qualified laboratory employees in cytology and/or pathology. The product is intended for professional use.

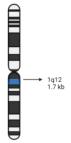
Clinical relevance

Within the chromosomes, the satellite DNA forms essential structures of chromosomes, i.e. telomeres and centromeres, ensuring protection and stability of chromatin of these sites. Satellites are highly variable in their sequence and in the number of copies both within a species and between closely related species that indicates rapid evolution of satellites. Satellite repeats are mostly located in centromeric and pericentromeric regions of chromosomes (Shatskikh et al., 2020). Errors within the pathways that regulate the assembly of the centromeres, can result in mis-segregation and aneuploidy of the chromosomal copy numbers. These are features for cancerous cells. Besides this, it can also result in chromosomal instability and, eventually chromosomal breakage (Barra & Fachinetti. 2018).

Probe specification

The CEP1-ISH probe consists of a 1.7 kb probe and is available in biotin and digoxigenin conjugated. CEP1-ISH probe is able to bind to the satellite repeats in the 1q12 region heterochromatin region of chromosome 1. The CEP1-ISH probes are pre-mixed in a hybridization mixture (formamide, dextran sulphate and SSC) and are ready to use solutions





Test principle

In an *in situ* hybridization assay, a double stranded DNA probe labelled with a hapten is used. The labelled DNA probe is diluted in a hybridization mixture. The hybridization mixture containing the DNA probe is added to the specimen. A co-denaturation will ensure that the genomic DNA and the probe DNA become single stranded. After denaturation, the probe DNA is able to hybridize to its complementary target sequences in the cells. In the REMBRANDT® CEP1-ISH assay, the haptens are attached to the probe and the signals can be visualized after detection by corresponding antibodies by fluorescent or brightfield microscopy.

Reagents provided

Product name Labelled CEP probe (Product number depending on label and	Amount size choice)
CEP 1-ISH probe biotin detection	C701P.0100.05	∑ 5 T
◆CEP 1-ISH probe biotin detection	C701P.0100.10	∑ 10 T
CEP1-ISH probe digoxigenin detection	C701P.9900.05	∑ 5 T
CEP1-ISH probe digoxigenin detection	C701P.9900.10	∑ 10 T
Pepsin powder	R011R.0000	1 g
Pepsin diluent	R018R.0000	15 ml
PanWash 4, 25X SSC	R025R.0000	15 ml

Assay procedure

REMBRANDT® CEP1-ISH detection assay procedure for cytological specimen.

- I. Incubate slides in pre-heated proteolytic work solution (prepare according to section 1.9 of Manual-FISH) (R011R.000 + R018R.000) at 37 °C (100 µg/ml) for 15 minutes followed by a brief rinsing in 0.01M HCI (1x 2 minutes) and subsequent rinses in PBS (2x 1 minute)
- II. Flush wash slides in deionised water, followed by dehydration in graded ethanol series (ethanol 70%, 96%, 96%, 100%, 100%) 1 minute each and air-dry slides for 15 minutes

Do not treat more than 5 slides at the same time, because the temperature of the pre-heated solutions may drop dramatically, thus causing incomplete pre-treatment. Additionally, allow the slides to air-dry as recommended; otherwise sections will be lost.

III. Homogenize probe solution (C701P.XXXX.YY) and spin briefly. Apply 15 μl of probe solution to each specimen. Cover all specimens with a cover slip (avoid air bubbles). Denature slides on a 80 °C hotplate or other heating device for 3 minutes

Work in a pre-set order to ensure that all slides have been incubated at 80 °C for the exact same time. Do not denature more than 5 slides at the same time, the temperature of the heating device may drop dramatically, thus causing incomplete denaturation.

- IV. Transfer the slides into a moist environment and incubate for 16 hours at 37 °C.
- Remove coverslips by soaking the slides in PBS at room temperature
- VI. Incubate the slides in diluted, pre-heated PanWash 4 (R025R.000) (prepare according to section 1.9 of Manual-FISH)

Do not incubate more than 5 slides at the same time in PanWash 4, the temperature of the PanWash 4 may drop dramatically, causing wrong stringency conditions.

VII. Appropriate detection system should be evaluated by the end-user. Recommended detection systems are listed below

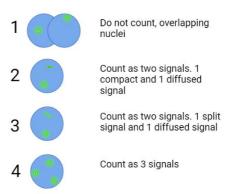
Digoxigenin detection	Biotin detection
R003R.0000	R041R.0000
Sheep aDig-AP conjugate	Goat aBio-AP Fab conjugate
R004R.0000 Sheep aDig-HRP conjugate	R042R.0000 Goat aBio-HRP Fab conjugate

Interpretation of results

The enumeration of chromosome 1 is conducted by microscopic examination of interphase nuclei (fluorescence or brightfield, depending on antibodies used for detection). The fluorescently or chromogenic-stained centromeres of chromosome 1 stand out brightly against the nucleus. The CEP1 procedure enables visual enumeration e.g. monosomy disomy, or trisomy of the centromeric region of chromosome 1 within the nuclei. The assay results are reported as the percentage of nuclei with 0, 1, 2, 3, 4, and >4 signals. Each signal corresponds to the centromere of chromosome 1

Enumerate the signals in the interphase nucleus using a 40X or 63X magnification. Objectives with higher magnification (eg, 63X or 100X) should be used to verify or resolve questions about split or diffused signals. Enumerate at least 100 nuclei per slide for accurate analysis.

- •Two signals in close proximity and approximately the same sizes but not connected by a visible link are counted as 2 signals.
- •Count a diffuse signal as 1 signal if diffusion of the signal is contiguous and within an acceptable boundary.
- •Two small signals connected by a visible link are counted as 1 signal.
- •Enumerate the number of nuclei with 0, 1, 2, 3, 4, or >4 signals. Count nuclei with 0 signals only if there are other nuclei with at least 1 signal present in the field of view. If the accuracy of the enumeration is in doubt, repeat the enumeration in another area of the slide.
- •Do not enumerate nuclei with uncertain signals



Performance characteristics

The REMBRANDT® CEP1-assay was analytically validated for CEP1-ISH green or orange detection. The results of the direct fluorescent assay are shown. However, for the CEP1-ISH assay, the detection system

may influence the performance characteristics and the CEP1-ISH in combination with different detection systems should be evaluated carefully by the end-user.

Analytical sensitivity

The analytical sensitivity was determined in three levels. The normal cut-off percentage was determined based on assessment of 200 individual nuclei in two samples of healthy interphase lymphocytes from peripheral blood. The beta inversion was used to determined the percentage of normal-cut off. For the noise-to-signal percentage, the noise and signal values were determined for 100 signals in interphase lymphocytes from peripheral blood of two independent samples. The differences between noise and signal were evaluated using a 95% confidence interval. For the hybridization efficiency, 200 individual nuclei were assessed for the presence of ISH signals.

Performance characteristic	Outcome
Normal cut-off percentage	8.6%
Noise-to-signal cut-off	4.8%
percentage Hybridization efficiency	100%

Analytical specificity

The analytical specificity was determined in two levels. The theoretical specificity was determined by sequencing analysis of the probe DNA and mapping on Hg38. The practical specificity was determined by assessing the hybridization pattern in metaphase chromosomes of interphase lymphocytes from peripheral blood.

Performance characteristic	Outcome
Theoretical specificity	Mapped on chromosome 1, g12
Practical specificity	100%

Limitations of Procedure

- The CEP1-ISH assay is solely applicable for the detection of the centromeric region of the human chromosome 1, which may be present in cell preparations (cytological specimen i.e. interphase lymphocytes from peripheral blood samples).
- ii) Either human tissue sections or human cytological preparations may be used. Samples must be fixed in buffered formalin or alcohol. In tissue sections are required, the sections should be prepared in a 4 µm thickness. Furthermore, the tissues should be glued to the glass slides with a bio-adhesive (e.g. organ silane), dried at room temperature, subsequently dried at 37 °C overnight and lastly completely deparaffinized in xylene and alcohol series and air dried
- iii) Cytological specimen should be prepared as required by the user, fixed with cytological fixation agent, rinsed in distilled water prior to the ISH procedure and air dried

- iv) Many factors can influence the performance of the ISH procedure. Failure in detection can be due to i.e. improper sampling, handling, the time lapse between tissue removal and fixation, the size of the tissue specimen in the fixation medium, the fixation time, processing fixed tissue, the thickness of the section, the bio-adhesive on the slide, deparaffinisation procedure, incubation times, proteolytic pre-treatment, detection reagents, incubation temperatures and interpretation of results.
- v) The performance of the ISH procedure is also affected by the sensitivity of the method and the presence of the centromeric region of chromosome 1. In case the limit of the sensitivity is reached a false negative reaction may be the result.
- vi) The CEP1-ISH assay results should not be relied on in case the sampling, sampling method, sample quality, sample preparation, reagents used, controls and procedure followed are not optimal or as described the working protocols.
- vii) The medical profession should be aware of risks and factors influencing the signal intensity while interpretating the test result. Microscopy settings might influence the signal intensity and/or interpretation.
- viii) Laboratory personnel performing the test should be trained and knowledgeable to be able to interpret the test results

Storage and handling

Store kit and its contents at 2-8°C. Store the dissolved and aliquoted reagents at recommended temperatures. When used and stored as indicated, the kit is stable until the expiry date printed on the box.

Product	Product number	Storage conditions
CEP1 Probe PanWash 4, 25X SSC	C701P.XXXX R025R.0000	2-8 °C Concentrated solution and diluted: 2-25 °C, ambient temperature
Pepsin powder	R011R.0000	Powder: 2-25°C, ambient temperature
Pepsin diluent	R018R.0000	Dissolved: -20°C Concentrated solution and diluted: 2-25°C, ambient temperature



Hazard statements

H315 - Causes skin irritation

H319 - Causes serious eve irritation

H351 - Suspected of causing cancer

H360D - May damage the unborn child

H373 - May cause damage to organs through prolonged or

repeated exposure

Precautionary Statements

P202 - Do not handle until all safety precautions have been read and understood

P280 - Wear protective gloves/protective clothing/eye protection/face protection

P302 + P352 - IF ON SKIN: Wash with plenty of water and soap P305 + P351 + P338 - IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing

P308 + P313 - IF exposed or concerned: Get medical advice/attention

P362 + P364 - Take off contaminated clothing and wash it before reuse

P405 - Store locked up

Additional information

Product in combination with other devices

The REMBRANDT® DNA probes are intended for standalone usage. The in vitro diagnostic is intended to be used in combination with standard cytological preparation methods, hot plate(s), stove(s), incubation device(s), water bath(s), temperature and incubation time control(s), proteolytic-, detection- and other reagents (not supplied with this reagent) and a microscope. The combination has been tested and validated. Since the standard formalin fixed, paraffin embedded tissue blocks, standard tissue freezing, tissue sectioning (microtome), standard cytological preparation methods, hot plate(s), stove(s), incubation device(s), water bath(s), temperature controls, incubation time control(s) and other not supplied reagents such as but not limited to proteolytic reagents, detection reagents and a microscope is not combined with the device as a product, conformity with the essential requirements is not applicable. Assay validation criteria are mentioned in 'Interpretation of the Results' and are also depending on the target load, which may influence the validation criteria.

For additional information regarding the REMBRANDT® assays, a manual is included which specifies the following subjects:

- Controls
- Materials required but not included
- Storage and shelf-life
- Performance precautions
- Preparations of reagents
- Specimen collection

- Quality control
- Trouble shooting guide

Technical assistance

For technical assistance regarding the products performance, please contact info@panpath.nl or call +31 495499090. Visit our website for reprints of datasheets or additional documentation. www.panpath.nl

Literature list

Barra, V., & Fachinetti, D. (2018). The dark side of centromeres: types, causes and consequences of structural abnormalities implicating centromeric DNA. Nature Communications, 9(1). https://doi.org/10.1038/s41467-018-06545-y

Shatskikh, A. S., Kotov, A. A., Adashev, V. E., Bazylev, S. S., & Olenina, L. V. (2020). Functional Significance of Satellite DNAs: Insights From Drosophila. Frontiers in Cell and Developmental Biology, 8(May), 1–19. https://doi.org/10.3389/fcell.2020.00312